

INSTITUTIONAL BIOSAFETY COMMITTEE (IBC)

Meeting Minutes

Date: Thursday, October 23, 2025

Time: 10:05 AM – 11:00 AM

Location: In-Person at Cold Spring Harbor Laboratory, Wendt CR, Cold Spring Harbor, NY

Members Present:

1. Sydney Gary, Research Operations (*IBC Chairperson*)
2. Lynn Blake, Research Compliance Officer
3. John Pisciotta, Environmental Health & Safety (*Biological Officer*)
4. Denise Roberts, Core Operations
5. Julie Cheong, Assistant Laboratory Building Manager
6. Scott Lyons, Animal Imaging (*Scientific Technical Advisor*)
7. Lisa Bianco, Laboratory Animal Resources (LAR) (*Technical Operations*)
8. Kenneth Addison, Clinical and Translational Collaborations
9. Kristen Panella (*Community Member, non-CSHL Affiliated*)

Members Not Present:

1. David Jackson, Plant Biology (*Plant Scientific Advisor*)
2. Janeen Braynen, Computational (*Plant Scientific Advisor*)
3. Rachel Rubino, Laboratory Animal Resources (LAR) (*Attending Veterinarian*)
4. Brian Kelleher (*Community Member, non-CSHL Affiliated*)

Non-Voting Attendees Present:

1. Joanie O'Connor, Administrative & Regulatory Coordinator
2. Angelina Regua, Research Compliance Officer (IRB)
3. Julie Sutherland, Conflict of Interest and Compliance Coordinator
4. Xiomara Medina, IACUC Administrator/Manager

Commonly Used Abbreviations

AAV: Adeno-associated viral vector

BSL: Biosafety level

BSO: Biosafety Officer

CRISPR: Clustered regularly interspaced short palindromic repeats

DMR: Designated Member Review

DURC: Dual Use Research of Concern

FCR: Full Committee Review

IACUC: Institutional Animal Care and Use Committee

IBC: Institutional Biosafety Committee

IRB: Institutional Review Board

NIH: National Institutes of Health

PI: Principal Investigator

PPE: Personal protective equipment

r/s NAM: Recombinant or synthetic nucleic acid molecules

RG: Risk Group

SOP: Standard Operating Procedures

Source material: blood, tissue, cell lines

1 CALL TO ORDER: The Institutional Biosafety Committee (IBC) Chairperson called the meeting to order at 10:05 AM. A quorum was present.

2 APPROVAL OF MINUTES:

The IBC Chair requested a motion to approve the minutes from the July 24, 2025, meeting and the September 26, 2025, ad hoc meeting. No questions or concerns were raised. A motion was made and seconded; the minutes were unanimously approved.

3 GENERAL OR OLD BUSINESS:

3.1 **Safe-tober Escape Room: Biosafety Awareness Day Update**
Research Compliance and Environmental Health & Safety (EH&S) have partnered to launch the inaugural institution-wide event, the "Safe-tober Escape Room: Biosafety Awareness Day." This collaboration invites all laboratory personnel to participate in engaging challenges to test biosafety knowledge and increase awareness. The event features an interactive escape room format with various games, promising a fun and educational experience for everyone.

4 OTHER BUSINESS:

4.1 **NIH IBC-RMS Submission Status**

The IBC-RMS submission was approved on September 3, 2025, with the next report due by September 3, 2026.

5 INDIVIDUAL PROJECT REVIEWS (FCR):

5.1 Arkarup Bandyopadhyay, 5-Year Renewal, IBC-2025-12, *Neural circuits for vocal communication in the singing mouse*

- NIH Guidelines Section III-D
- The Bandyopadhyay lab uses singing mice as a model to investigate the neural circuits in the brain that underlie vocal communication in mammals.
- This project has an approved IACUC protocol number 2023-1330.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
- PPE will be worn as dictated by the biosafety level of the research.
- The Committee unanimously approved the IBC submission and use of agents as described, with a vote of 9-0-0 (for, against, abstained).

BSL-1 Injection of AAV (no helper virus), including CRISPR, into whole animals.

BSL-2 Injection of Sindbis into whole animals.

BSL-2 Injection of pseudorabies into whole animals.

BSL-2+ Use of flow cytometry.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

5.2 David Jackson, 5-Year Renewal, IBC-2025-10, *Investigation of molecular mechanisms controlling plant development*

- NIH Guidelines Section III-D-5

- The Jackson lab studies genes and signals in cells that regulate the growth and shape of plants.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
- PPE will be worn as dictated by the biosafety level of the research.
- The Committee unanimously approved the IBC submission and use of agents as described, with a vote of 9-0-0 (for, against, abstained).

BSL-1 Transgenic plant work with Arabidopsis and maize.

BSL-1 CRISPR-Cas9 mutagenesis of genes in maize and Arabidopsis.

BSL-1 Use of protein expressions in plants (Agrobacterium and tobacco leaves) for protein production.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

5.3 Zachary Lippman, 5-Year Renewal, IBC-2025-11, *Plant and Crop Development, Genomics and Genetics*

- NIH Guidelines Section III-D-5
- The Lippman lab integrates development, genetics, genomics, and genome editing to uncover, explore, and exploit the mechanisms that determine how plant stem cells become shoots and flowers.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
- PPE will be worn as dictated by the biosafety level of the research.
- The Committee unanimously approved the IBC submission and use of agents as described, with a vote of 9-0-0 (for, against, abstained).

BSL-1 Transgenic plant work with tomatoes, close crop relatives of tomato from the Solanaceae family, primarily eggplant and related species (*Solanum prinophyllum*, *Solanum aethiopicum*, *Solanum anguivi*, *Solanum sisymbriifolium*, *Solanum cleistogamum*, *Solanum macrocarpon*, *Solanum atropurpureum*, *Solanum citrullifolium*, *Solanum rostratum*, *Solanum dasyphyllum*, *Physalis grisea*, *Physalis peruviana*) and Arabidopsis.

BS-1 Use of deactivated Tobacco Rattle Virus to deliver guide RNAs to Cas9-expressing plants.

BSL-1 Use of CRISPR.

BSL-2+ Use of flow cytometry.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

5.4 Semir Beyaz, amendment, IBC-2023-7, *Metabolic and Epigenetic Regulation of Tissues Regeneration, Immunity and Cancer*

- NIH Guidelines Section III-D

- The Beyaz lab deciphers how nutritional cues and metabolic-epigenetic crosstalk sculpt cellular networks that sustain healthy tissue function-and how their disruption drives maladaptive remodeling in diseases such as colon and endometrial cancers, as well as in endometriosis.
- This project has an approved IACUC protocol number 2021-1159.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
- PPE will be worn as dictated by the biosafety level of the research.
- The Committee unanimously approved the IBC amendment with a 9-0-0 (for, against, abstained), pending a support/approval letter from Rockefeller, who is providing the parasite for these experiments.

BSL-2 Use of *Strongyloides venezuelensis* to infect animal models.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved effective October 23, 2025. The approval letter was generated on December 16, 2025, after the required information was received and approved.

5.5 Tobias Janowitz, amendment, IBC-2024-7, *Immunological, dietary, endocrinological, physiological and pharmacological regulation of anti-cancer immunity*

- NIH Guidelines Section III-D
- The Janowitz lab investigates the connections between metabolism, endocrinology, and immunology to discover how the body's response to a tumor can be used to improve treatment for patients with cancer.
- This project has an approved IACUC protocol number 2023-1329.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
- PPE will be worn as dictated by the biosafety level of the research.
- The Committee unanimously approved the IBC amendment with a 9-0-0 (for, against, abstained).

BSL-2 Use of Diphtheria Toxin (DT) for in vivo animal injections.

BSL-2 Use of Diphtheria Toxin (DT) for cell treatment.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

5.6 Leemor Joshua-Tor, amendment, IBC-2024-5, *Nucleic-Acid Regulatory Pathways*

- NIH Guidelines Section III-D
- The Joshua-Tor lab studies the molecular basis of nucleic acid regulatory processes using tools of structural biology and biochemistry.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.

- PPE will be worn as dictated by the biosafety level of the research.
- The Committee unanimously approved the IBC amendment with a 9-0-0 (for, against, abstained).

BSL-2 Culturing and use of non-viral transfection of human cell lines HEK (Flp-In 293™ T-Rex cell line) and HeLa (FLP-In TReX HeLa) to isolate and sequence their RNA under different conditions.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

5.7 Michael Lukey, amendment, IBC-2025-3, *Metabolic reprogramming during tumorigenesis and metastasis*

- NIH Guidelines Section III-D
- The Lukey lab studies the nature and regulation of metabolic adaptation during tumorigenesis and metastasis, with the intention of identifying metabolic vulnerabilities that can be targeted for cancer therapy.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
- PPE will be worn as dictated by the biosafety level of the research.
- The Committee unanimously approved the IBC amendment with a 9-0-0 (for, against, abstained).

BSL-1 Protein expression for purification using insect cells expressing metabolic enzymes from pFL vector assembled into bacmids.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

6 DESIGNATED MEMBER REVIEW (DMR):

6.1 Ivan Iossifov, annual review, IBC-2022-11, *Development of Genomics and System Biology Methods for the Study of the Genetic Architecture of Autism and Related Neurodevelopmental Disorders*

- NIH Guidelines Section III-F
- The Iossifov lab studies the genetics of common diseases in humans using two main tools: next-generation sequencing and molecular networks representing functional relationships among genetic loci.
- This project does not involve recombinant or synthetic nucleic acid molecules (r/s NAM); it can proceed without requiring specific biosafety containment.
- The protocol was approved, effective October 23, 2025.

6.2 David Klindt, annual review, IBC-2024-17, *Finding Interpretable Representations in Neural Networks*

- NIH Guidelines Section III-F
- The Klindt lab studies the intersection of biological systems and artificial intelligence (AI), aiming to understand how organisms, particularly the brain, process sensory information and generalize knowledge across different contexts.

- This project does not involve recombinant or synthetic nucleic acid molecules (r/s NAM); it can proceed without requiring specific biosafety containment.
 - The protocol was approved, effective October 23, 2025.
- 6.3 Peter Koo, annual review, IBC-2024-10, *Utilizing Deep Learning to Elucidate Biological Mechanisms*
- NIH Guidelines Section III-F
 - The Koo lab develops methods to interpret high-performing deep learning models to distill knowledge that they learn from big, noisy, biological sequence data. Their goal is to elucidate biological mechanisms that underlie sequence-function relationships for gene regulation and protein (dys)function.
 - This project does not involve recombinant or synthetic nucleic acid molecules (r/s NAM); it can proceed without requiring specific biosafety containment.
 - The protocol was approved, effective October 23, 2025.
- 6.4 Alexander Krasnitz, annual review, IBC-2022-7, *Computational Biology of Human Diseases*
- NIH Guidelines Section III-F
 - The Krasnitz lab develops mathematical and statistical tools to investigate the population structure of cells comprising a malignant tumor and to reconstruct evolutionary processes leading up to that structure.
 - This project does not involve recombinant or synthetic nucleic acid molecules (r/s NAM); it can proceed without requiring specific biosafety containment.
 - The protocol was approved, effective October 23, 2025.
- 6.5 Adam Siepel, annual review, IBC-2024-14, *Evolutionary Human Genomics*
- NIH Guidelines Section III-F
 - The Siepel lab focuses on questions involving molecular evolution and transcriptional regulation, with applications to cancer and other diseases as well as to plant breeding and agriculture.
 - This project does not involve recombinant or synthetic nucleic acid molecules (r/s NAM); it can proceed without requiring specific biosafety containment.
 - The protocol was approved, effective October 23, 2025.
- 6.6 Katherine Alexander, annual review-no changes, IBC-2024-18, *Principles of Speckle-based Gene Regulations*
- NIH Guidelines Section III-D
 - The Alexander lab focuses on the fundamental mechanisms of gene regulation as mediated by nuclear speckles.
 - A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
 - PPE will be worn as dictated by the biosafety level of the research.
 - In the annual review, which resulted in no protocol changes, the Committee unanimously approved the IBC submission and use of agents as described, with a vote of 9-0-0 (for, against, abstained).

BSL-1 Standard subcloning and bacterial expression of plasmids; Routine plasmid work; Subcloning and non-pathogenic E. Coli culture for plasmid expression and amplification to be used for cell line transfections; Use of mammalian (non-viral) materials (tissue culture); Use of non-mammalian vectors (luciferase reporter plasmids) and bacterial hosts (DH5 α cells); Standard subcloning and bacterial expression with the addition of bacterial vectors, plasmids, and hosts.

BSL-1 Non-viral mammalian expression.

BSL-2+ Use of amphotropic lentivirus, including CRISPR, to test transcription factor DNA-speckle targeting or examine nuclear speckle functions.

BSL-2+ Use of neuroblastoma, cancer, and mouse cell lines.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

6.7 Doug Fearon, annual review-no changes, IBC-2024-11, *Immunotherapy of mouse pancreatic cancer with anti-hCXCR4*

- NIH Guidelines Section III-D
- The Fearon lab studies the interaction between cancer and the immune system. Their underlying premise is that the tumor microenvironment is immune suppressive because cancer cells elicit responses characteristic of wound healing and tissue regeneration.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
- PPE will be worn as dictated by the biosafety level of the research.
- In the annual review, which resulted in no protocol changes, the Committee unanimously approved the IBC submission and use of agents as described, with a vote of 9-0-0 (for, against, abstained).

BSL-1 Lentiviral vectors and genes are cloned and expanded typically in E. coli.

BSL-1 Standard subcloning, bacterial expression, and non-viral mammalian expression.

BSL-2 Use of ecotropic lentivirus in human and mouse cell lines to induce overexpression or knock out of CXCR4, TNFRSF1B, and TNFRSF1A.

BSL-2 Use of ecotropic adenovirus in the mouse cell line to flip out GFP gene in transduced cell lines.

BSL-2+ Use of flow cytometry.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

6.8 Thomas Gingeras, annual review-no changes, IBC-2024-12, *Characterization of Extracellular Vesicles*

- NIH Guidelines Section III-D
- The Gingeras lab studies where and how functional information is stored in genomes. These efforts help explain the biological and clinical effects of disease-causing gene mutations in humans and other organisms.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications,

recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.

- PPE will be worn as dictated by the biosafety level of the research.
- In the annual review, which resulted in no protocol changes, the Committee unanimously approved the IBC submission and use of agents as described, with a vote of 9-0-0 (for, against, abstained).

BSL-1 Genome, epigenome, and transcriptional analyses on engineered cells.

BSL-2 Isolation of genomic DNA and RNA of B cells.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

6.9 Helen Hou, annual review-no changes, IBC-2022-16, *Neurobiological Control of Facial Expression in Health and Disease*

- NIH Guidelines Section III-D
- The Hou lab studies how the brain orchestrates motor control in natural and innate behaviors, specifically the neural mechanisms underlying the dynamic control of facial expression using rodents as a model, applying electrophysiological, imaging, neuroanatomical, behavioral, and computational analyses.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
- PPE will be worn as dictated by the biosafety level of the research.
- In the annual review, which resulted in no protocol changes, the Committee unanimously approved the IBC submission and use of agents as described, with a vote of 9-0-0 (for, against, abstained).

BSL-1 Stereotactic brain injection of rAAVs (ecotropic, non-replicating) to visualize anatomical connections as well as monitor and perturb activity in the nervous system.

BSL-2 Stereotactic brain injection of modified pseudotyped G-deleted rabies (ecotropic, non-replicating, no propagation) to visualize anatomical connections.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

6.10 Scott Lyons, annual review-no changes, IBC-2024-13, *Lentiviral and transposon-based vectors for the introduction of stable transgene expression in vitro, ex vivo, and in vivo*

- NIH Guidelines Section III-D
- The Lyons lab supports researchers in the area of preclinical in vivo imaging. They also develop new and innovative ways to image tumors, such as their cellular proliferation or metastatic spread, with the ultimate goal of discovering effective treatments and better understanding their mechanism of action.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have

been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.

- PPE will be worn as dictated by the biosafety level of the research.
- In the annual review, which resulted in no protocol changes, the Committee unanimously approved the IBC submission and use of agents as described, with a vote of 8-0-0 (for, against, abstained).

BSL-1 Routine plasmid work.

BSL-1 Work with transposon-based vectors (non-viral).

BSL-1 Use of CRISPR/Cas9 to image tumor-related biology (non-viral).

BSL-2 Routine tissue culture, lentiviral and adenoviral vector work (transgenic contents of these vectors will not contain transgenic material that is cell-transforming or that can affect gene expression in human cells).

BSL-2 Use of new variant reporter transgene classes.

BSL-2+ All lentiviral vector work containing transgenic material (e.g., shRNA or gRNA) that can modify the expression of genes in human cells.

BSL-2+ Use of flow cytometry.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

6.11 Alea Mills, annual review-no changes, IBC-2022-18, *Genetic and Epigenetic Basis of Cancer*

- NIH Guidelines Section III-D
- The Mills lab studies genetic pathways important in cancer, aging, and autism, identifying the genetic players and determining how aberrations in their functions culminate in human disease.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
- PPE will be worn as dictated by the biosafety level of the research.
- In the annual review, which resulted in no protocol changes, the Committee unanimously approved the IBC submission and use of agents as described, with a vote of 9-0-0 (for, against, abstained).

BSL-1 Use of PCR and restriction digests to subclone and express in bacterial systems.

BSL-2 Perform transient transfections with the aid of lipids such as PEI and Lipofectamine in cell lines (no virus used, only plasmid).

BSL-2+ Work with amphotropic lentivirus and amphotropic retrovirus expression vectors, including CRISPR, to knock out genes in various cell lines.

- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

6.12 Ullas Pedmale, annual review-no changes, IBC-2021-05, *Research on Arabidopsis cryptochromes*

- NIH Guidelines Section III-D-5
- The Pedmale lab studies how the environment of a plant modulates its growth and development. Understanding environmental control of growth will have far-

reaching implications for agriculture, energy production, and many other human activities.

- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
- PPE will be worn as dictated by the biosafety level of the research.
- In the annual review, which resulted in no protocol changes, the Committee unanimously approved the IBC submission and use of agents as described, with a vote of 9-0-0 (for, against, abstained).
 - BSL-1 Subcloning, bacterial expression, and yeast two-hybrid.
 - BSL-1 Cloning for the expression of genes in bacteria and plants.
 - BSL-1 Genotyping of mutants and transgenic plants.
- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

6.13 Jessica Tollkuhn, annual review-no changes, IBC-2024-15, *Hormonal regulation of gene expression in the brain*

- NIH Guidelines Section III-D
- The Tollkuhn lab studies how estrogen and testosterone regulate gene expression in the brain. The receptors for these steroid hormones directly bind DNA to turn genes on or off.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
- PPE will be worn as dictated by the biosafety level of the research.
- In the annual review, which resulted in no protocol changes, the Committee unanimously approved the IBC submission and use of agents as described, with a vote of 9-0-0 (for, against, abstained).
 - BSL-1 Use of ecotropic, replication-incompetent AAV to infect animal models.
 - BSL-1 Transcardiac perfusion of injected mice.
 - BSL-1 Use of mammalian (non-viral) materials (tissue culture), including mouse immortalized hypothalamic cell lines.
 - BSL-1 Standard rDNA protocols and subcloning in bacterial, production, and storage of helper-free AAV.
 - BSL-1 Use of non-mammalian vectors and bacterial hosts (DH5α cells).
 - BSL-2+ Stereotactic brain injection of modified pseudotyped G-deleted Rabies (rabies).
 - BSL-2+ Use of flow cytometry.
- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

6.14 Michael Wigler, annual review-no changes, IBC-2022-20, *Wigler IBC Renewal*

- NIH Guidelines Section III-D

- The Wigler lab studies human cancer and the contribution of new mutations to genetic disorders.
- A proper risk assessment has been performed, and the following applicable agents, their characteristics and sources, hosts, modifications, recommended biosafety containment levels, and other pertinent details have been reviewed for the potential risks associated with the protocol to ensure a safe and responsible research environment.
- PPE will be worn as dictated by the biosafety level of the research.
- In the annual review, which resulted in no protocol changes, the Committee unanimously approved the IBC submission and use of agents as described, with a vote of 9-0-0 (for, against, abstained).
 - BSL-1 Isolation of human DNA, RNA, and PCR.
 - BSL-1 Transient transfection of human cultured cells with Baculovirus-containing GFP/RFP constructs.
 - BSL-2 Routine cell culture of human cell lines.
 - BSL-2 Preparation of human tissues and fluids for isolation of DNA/RNA or flow cytometry.
 - BSL-2 Use of flow cytometry for sorting isolated nuclei from mouse and human cells.
 - BSL-2+ Use of flow cytometry for sorting human cell cultures and clinical specimen samples.
- Training: All personnel have completed the required biological training in accordance with institutional and NIH guidelines.
- The protocol was approved, effective October 23, 2025.

7 CONTINUING EDUCATION

7.1 Biosafety Awareness Day: Jeopardy Game

As part of ongoing education and the upcoming biosafety awareness day, the DIY Jeopardy board game was shown to the committee as an example of an event activity. Members of the committee played the game and assessed their level of expertise.

8 PUBLIC COMMENTS

- 8.1 For security reasons, Cold Spring Harbor Laboratory has an institutional policy that information regarding IBC meeting times and locations is not publicly available. Any member of the public requesting IBC information is kindly requested to contact the CSHL Office of Research Compliance at ResearchCompliance@cshl.edu.

9 MEETING ADJOURNED AT APPROXIMATELY 11:00 AM